

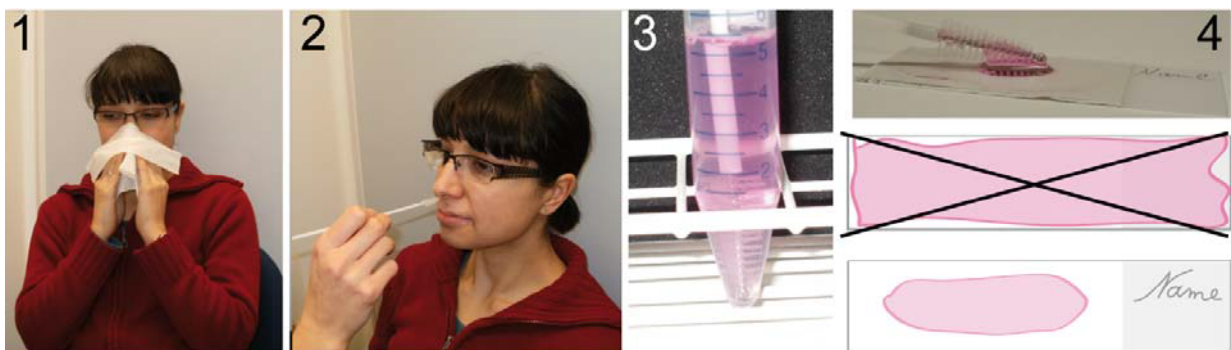
Nasal brushing Protocol

Procedure:

- Heat the medium to room temperature. Wet the brush with sterile isotonic saline solution. We use Celletta™ brush cell collector with protective tip (product number 9100060; Engelbrecht Medizin-und Labortechnik GmbH, Tiefenbachweg 13; 34295 Edermünde, Germany). Other brushes can also be used.
- Ask the patient to clean his nose. The patient should sit in a chair with the head against a wall so he can't move the head backwards (figure 1). Alternatively a 2nd person should stand behind him and hold the head. Ask the patient to hold the collecting tube so his hands are occupied with this.
- Examine the inferior nasal meatus with the rhinoscope (if possible), the mucosa should look clean and healthy.
- Take the brush and rub it a few times rapidly against the medial and superior side of the inferior nasal meatus, using rotatory and linear movements (figure 2). The patient feels a local burning and will probably get tears in the eye but the discomfort will diminish rapidly.
- Take the brush out and put it immediately in the collecting tube (figure 3). Shake the brush vigorously within the tube at least 40 times, so that the cells become detached from the brush.

Preparing slides for Immuncytochemistry

- Please include data on direct microscopy (Ciliary function) and electron microscopy findings if available
- Use pre-cleaned slides without additional preparations (ordinary slides). Take the brush to portion out the suspension onto slides (see figure 4). Please prepare ~25 slides until **no fluid** is left in the tube. Let the slides dry completely at room temperature. This will take a few hours. Label the slides with a pencil and send them to our lab address:



Labor Univ.-Prof. Dr. med. Heymut Omran
Universitätsklinikum Münster, Klinik für Kinder- und Jugendmedizin
-Allgemeine Pädiatrie-
Albert-Schweitzer-Campus 1, Gebäude: A1
48149 Münster

Tel: +49-(0)251-83-40011 Fax: +49-(0)251-83-40010, E-mail: Labor-Omran@ukmuenster.de